

Evaluation of antioxidant potential and antibacterial activity of *Calotropis gigantea* and *Vinca rosea* using *invitro* model

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Abstract

The different parts of *Calotropis gigantea* and *Vinca rosea* belonging to the families of *Asclepiadaceae* and *Apocynaceae* were studied for their antioxidant and antimicrobial activities against selected bacterial strains. From the results it was evident that the flower of *Vinca rosea* showed the highest antioxidant activity of 97.44% at 800 µg which was higher than the standard L-ascorbic acid (94%) and *Calotropis gigantea* showed the least. *Calotropis gigantea* exhibited highest inhibition zone of 14, 14 and 11 mm against *Escherichia coli*, *Salmonella typhi* and *Shigella sonnei* when compared to *Vinca rosea*. Hence, the present study supports the view, that these medicinal plants might be useful as antioxidant and antimicrobial agents.

Keywords: DPPH assay, antioxidant, antimicrobial, *Vinca rosea*, bacterial strains.

Introduction

Research in the chemistry of natural products has endless potential and is especially important in countries like India which has a rich biodiversity. In the recent years interest in the study of antioxidant activity of plant extracts and isolation from plants has grown due to the fact that the free radicals have been related to degenerative diseases (Willcox *et al.*, 2004). Human cells are constantly exposed to reactive oxygen radicals generated by a number of biotic and abiotic factors such as irradiation, environmental factors, pollutants, stress or byproducts of metabolic processes. When the exposure overwhelms endogenous preventive systems, cells are exposed to potentially harmful load of oxidants, leading to various free radicals induced noxious effects. Free radical attacks biological molecules such as lipids, proteins, enzymes, DNA and RNA leading to cell or tissue injury associated with many diseases including ageing, atherosclerosis, heart diseases and carcinogenesis (Halliwell, 1994). Antioxidants are compounds which act as radical scavengers when added to the food products and prevent the radical chain reaction of oxidation, delay or inhibit the oxidation process and increase shelf life by retarding the processes of lipid peroxidation (Young *et al.*, 2001). The ability of phenolic substances including flavonoids and phenolic acids acting as antioxidants has been reported (Liu *et al.*, 2003). Tannins have been reported to have strong antioxidant activity (Cai *et al.*, 2006). There is also growing interest both in industry and in scientific research in spices and medicinal herbs because of their antimicrobial and antioxidant activity (Eyob *et al.*, 2008).

Vinca rosea has a variety of medicinal properties such as antibacterial (Carew *et al.*, 1970), antifungal (Jaleel *et al.*, 2007), antiviral (Fransworth *et al.*, 1968), anticancer (Ram *et al.*, 2001). *Calotropis gigantea* has been reported to possess a number of medicinal properties and

is used in toothache, earache, sprain, anxiety, pain, epilepsy, mental disorder and also it possesses anti-diarrheal, analgesic and CNS activity (Pathak & Argal, 2007).

An extensive literature survey indicates antioxidant and antimicrobial activity in *Calotropis gigantea* and *Vinca rosea*. But only scanty information is available on such potential regarding the individual plant parts concerned (root, stem, leaf, flower & seed). In the present research study, the experiment was carried out in triplicate so as to compare the antioxidant and antimicrobial activities of the different parts of the two selected Indian medicinal plants and the values were compared with that of the previous reports. The concentration dependency of the antioxidant and antimicrobial activities was also investigated.

Materials and methods

All chemicals and solvents were of analytical grade (Sigma-Aldrich). The bacterial strains were purchased from the Dept. of Molecular Biology, Christian Medical College and Hospital, Vellore, Tamil Nadu, India. *Calotropis gigantea* and *Vinca rosea* were identified by Ms. Isabella Roseline, Head, Dept. of Botany in Auxilium College campus and authenticated by a Taxonomist Mr. Babu, Cholayil, Chennai and the Vouchers of the plant specimen were deposited in the Dept. of Botany, Auxilium College with the code DRC_cg1 and DRC_vr1.

Different parts of the plant materials like root, stem, leaves, flower and seeds were shade dried, finely ground and were percolated in 100% methanol and filtered. The filtrate was evaporated at 40°C under reduced pressure by a rotary evaporator to give a semisolid residue of approximately 140 g each.

Cultures of seven human pathogenic bacteria belonging to 2 gram positive and 5 gram negative bacteria obtained from the Dept. of Microbiology, Christian Medical College (CMC), Vellore were used for

the *invitro* antibacterial assay. The bacterial strains *Escherichia coli* (ATCC25922), *Klebsiella pneumonia* (ATCC10031), *Salmonella typhi* (ATCC10749), *Shigella sonnei* (ATCC25931), *Bacillus cereus* (ATCC10987), *Salmonella paratyphi* (ATCC11511) and *Staphylococcus aureus* (ATCC25923) were chosen for the present study.

Radical scavenging activity

Experiments were carried out in triplicate, according to the method of Blois (1958) with the slight modification in Cakir *et al.* (2003). Briefly, 25 mg/l solution of DPPH radical (Aldrich) in methanol was prepared and then 2 ml of this solution was mixed with different concentration (400, 600 & 800 µg) of sample solution to achieve the final volume of 3 ml. After 30 min the absorbance was measured at 517 nm. Decrease in the absorbance of the DPPH solution indicates an increase of the DPPH antioxidant activity. The antioxidant activity (AOA) was calculated using the equation:

$$\text{AOA} = \frac{\text{Ao} - \text{As}}{\text{Ao}} \times 100$$

Ao = DPPH solution without the sample

As = DPPH solution with the sample

Antimicrobial activity

Sterile nutrient broth was inoculated with freshly isolated bacterial culture and incubated for 24 h at 37°C. The bacterial suspension was found to be approximately 10^7 - 10^8 cells/ml. After the incubation period they were used as inoculum. About 0.1 ml of the suspension containing 10^8 colony forming unit (CFU/ml) of bacterial strains was taken to study by agar diffusion method (Estevinho *et al.*, 2008). About 500 mg/ml of methanolic extracts of the plant materials with different concentrations like 5, 10, 12.5 and 25 mg of sample were used and their zones of inhibitions were monitored after 24 h and the inhibition zone was compared with methanol which was a negative control.

Statistical analysis

The experiments were carried out in triplicate and the statistical software package (SPSS 12.0) was used for the statistical analysis and the results were given as a mean \pm standard deviation (SD). Regression analysis was carried out for the comparison of concentration dependency and one-way analysis of variance (ANOVA) was used for comparison of more than two means. A difference was considered statistically significant when $p \leq 0.05$ (Sabir & Rocha, 2008).

Results and discussion

Radical scavenging activity of *Calotropis gigantea* and *Vinca rosea*

The results of the antioxidant activity of *Calotropis gigantea* and *Vinca rosea* determined by DPPH assays at

different concentrations are given in Table 1 & 2. From Table 1 it was evident that all parts of *C. gigantea* showed moderate antioxidant activity when compared with standard antioxidant L-ascorbic acid whose antioxidant activity at different concentrations like 400, 600 and 800 µg were 84%, 88% and 94% and the antioxidant activity of L-ascorbic acid at 800 µL (positive control) were in good agreement with the results of earlier workers (Taechowisan *et al.*, 2009). The antioxidant activities followed the trend: stem 65.89% > flower 54.51% > root 44.29% > leaves 23.64%. The radical scavenging activities of flower showed similar antioxidant activity at different concentrations. This clearly indicates that the antioxidant activity was highest at the 400 µg and the increase in concentration showed least effect on the antioxidant activity. It was concentration independent as proved by regression analysis.

Table 1. Antioxidant potential of different parts of *Calotropis gigantea*.

Different parts	Conc. (µg)	Mean absorbance \pm SD	R ²	Antioxidant activity (%)
Leaves	400	0.3895 \pm 0.0057	0.955	15.14
	600	0.3627 \pm 0.0056		20.97
	800	0.3505 \pm 0.0049		23.64
Stem	400	0.2427 \pm 0.0065	0.937	31.43
	600	0.2092 \pm 0.0069		40.89
	800	0.1207 \pm 0.0065		65.89
Root	400	0.3110 \pm 0.0092	0.990	18.16
	600	0.2252 \pm 0.0074		30.90
	800	0.1025 \pm 0.0053		44.29
Flower	400	0.2622 \pm 0.0073	0.997	51.25
	600	0.2526 \pm 0.0067		52.37
	800	0.2447 \pm 0.0056		54.51

Table 2 shows the antioxidant activities of different parts of *V. rosea* at various concentrations. From the Table it was clear that all the parts of *V. rosea* showed the highest antioxidant activity and followed the trend flower (97.44%) > stem (93.80) = root (93.84) > leaves (83.72) > seed (80.28) and the antioxidant activity of flower was higher

Table 2. Antioxidant potential of different parts of *Vinca rosea*.

Different parts	Conc. (µg)	Mean absorbance \pm SD	R ²	Antioxidant activity (%)
Leaves	400	0.0652 \pm 0.0092	0.372	82.36
	600	0.0790 \pm 0.0076		82.56
	800	0.0737 \pm 0.0083		83.72
Stem	400	0.0520 \pm 0.0057	0.967	86.87
	600	0.0427 \pm 0.0046		89.20
	800	0.0247 \pm 0.0069		93.75
Root	400	0.0322 \pm 0.0074	0.797	93.15
	600	0.0292 \pm 0.0069		93.79
	800	0.0290 \pm 0.0076		93.84
Flower	400	0.1252 \pm 0.0079	0.979	54.29
	600	0.0812 \pm 0.0067		70.35
	800	0.0070 \pm 0.0071		97.44
Seed	400	0.2240 \pm 0.0087	0.950	52.24
	600	0.1845 \pm 0.0066		60.66
	800	0.0925 \pm 0.0083		80.28

than L-ascorbic acid at 800 µg. This may be due to the presence of phenolic compounds and flavonoids as detected by photochemical analysis which were responsible for the antioxidant activity even at lower concentrations (Villano *et al.*, 2007). The antioxidant activities of leaves, stem and root were independent of concentration which shows that it may contain phytochemicals that are responsible for antioxidant activities as it was evident from the Table 5. From the Table 1 & 2 it was also evident that good precision was observed which was evident from the standard deviation values.

Table 3. Antibacterial activity of *Calotropis gigantea*.

Plant parts	Conc. (mg)	<i>B.c</i> (mm)	<i>E.c</i> (mm)	<i>K.p</i> (mm)	<i>S.a</i> (mm)	<i>S.s</i> (mm)	<i>S.p</i> (mm)	<i>S.t</i> (mm)
Leaves	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	-	-	-	-	-	-	-
	25.0	-	8	9	8	-	-	6
Stem	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	-	-	-	-	-	-	-
	25.0	-	-	6	-	-	-	-
Root	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	6	-	-	-	6	6	-
	25.0	9	-	-	-	8	8	-
Flower	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	6	6	-	-	-	-	6
	25.0	9	14	6	6	11	9	14

Bacillus cereus (*B.c*); *Escherichia coli* (*E.c*); *Klebsiella pneumonia* (*K.p*); *Staphylococcus aureus* (*S.a*); *Shigella sonnei* (*S.s*); *Salmonella paratyphi* (*S.p*); *Salmonella typhi* (*S.t*)

Table 4. Antibacterial activity of *Vinca rosea*.

Plant parts	Conc. (mg)	<i>B.c</i> (mm)	<i>E.c</i> (mm)	<i>K.p</i> (mm)	<i>S.a</i> (mm)	<i>S.s</i> (mm)	<i>S.p</i> (mm)	<i>S.t</i> (mm)
Leaves	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	-	-	6	-	-	9	-
	25.0	-	-	8	-	-	11	-
stem	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	-	-	-	-	-	6	-
	25.0	-	6	-	7	-	10	6
Flower	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	-	8	-	-	6	6.5	-
	25.0	-	11	10	6	8	10	6
Root	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	-	-	6	6	-	-	-
	25.0	-	6	8	9	6	6	6
Seed	05.0	-	-	-	-	-	-	-
	10.0	-	-	-	-	-	-	-
	12.5	-	-	-	-	-	-	-
	25.0	6	6	-	-	-	-	6

Bacillus cereus (*B.c*); *Escherichia coli* (*E.c*); *Klebsiella pneumonia* (*K.p*); *Staphylococcus aureus* (*S.a*); *Shigella sonnei* (*S.s*); *Salmonella paratyphi* (*S.p*); *Salmonella typhi* (*S.t*)

From the regression analysis it was clear that there was a good linearity between concentration and absorbance for all the parts of *C. gigantea* and *V. rosea* except the root and leaves of *V. rosea* whose R^2 value were 0.797 and 0.372 respectively. Hence, the root and leaves of *V. rosea* were concentration independent at experimental concentrations. The poor linearity in leaves of *V. rosea* ($R^2 = 0.372$) may be due to the interfering plant pigments. One way analysis of variance (ANOVA) was used to test the level of significance between absorbance and concentration. The experiments were conducted in triplicate and the probability factor (P) was <0.05 between different parts of *C. gigantea* and *V. rosea* and so it was considered as significant.

Antibacterial activity

In the present study, the methanolic extracts of various parts of *C. gigantea* and *V. rosea* were selected for antibacterial activity on seven different microorganisms- *Escherichia coli* (*E.c*), *Klebsiella pneumonia* (*K.p*), *Salmonella typhi* (*S.t*), *Shigella sonnei* (*S.s*), *Bacillus cereus* (*B.c*), *Salmonella paratyphi* (*S.p*) and *Staphylococcus aureus* (*S.a*). Depending on the measured values of the complete inhibition diameter in mm, the antibacterial activity can be classified into the following types, such as >12 mm zone of inhibition-high sensitivity, 9-12 mm zone of inhibition-moderate sensitivity, 6-9 mm zone of inhibition-less sensitivity and <6 mm zone of inhibition-resistant (Uma Devi *et al.*, 2007).

Antibacterial activity of *Calotropis gigantea*.

The antibacterial activities of various parts of *C. gigantea* for selected test bacterial strains at different concentrations were given in Table 3. From the Table it was clear that, all the samples at different concentrations (5, 10, 12.5 & 25 mg) gave different inhibition activities towards tested organisms when compared with the negative control methanol whose inhibition zone was 6 mm. Among the different plant extracts, flower showed the highest antibacterial activity against *Escherichia coli* and *Salmonella typhi* with the net inhibition zone of 14 mm at 25 mg. Root exhibited moderate sensitivity against *Shigella sonnei*, *Bacillus cereus* and *Salmonella paratyphi*. The stem was bacterial resistant to all the selected bacterial strains.

Antibacterial activity of *Vinca rosea*.

Preliminary screening for antibacterial activities of different parts like leaves, stem, flower, root and seed of *V. rosea* against the seven selected bacterial strains at various concentrations are given in Table 4. The flower

Table 5. Phytochemical analysis of *Calotropis gigantea* & *Vinca rosea*.

Phytochemicals	<i>Calotropis gigantea</i>				<i>Vinca rosea</i>			
	Leaves	Root	Stem	Flower	Leaves	Root	Stem	Flower
Phenolic compounds	-	-	-	+	+	-	-	+
Steroids	+	+	+	+	-	+	+	-
Terpenoids	-	-	-	-	-	-	-	-
Flavonoids	-	-	-	+	+	-	-	+
Sugars	+	+	+	+	+	+	+	+
Coumarin	-	-	-	-	+	+	+	+
Quinone	-	-	-	+	+	-	+	+
Tannins	-	-	-	-	+	-	-	-
Saponin	+	+	+	+	+	+	+	+

+ indicates presence; - indicates absence.

exhibited different zone of inhibition for all tested bacterial strains- *Escherichia coli*, *Klebsiella pneumonia*, *Staphylococcus aureus*, *Shigella sonnei*, *Salmonella paratyphi* and *Salmonella typhi* except *Bacillus cereus*. Leaves and root showed moderate sensitivity against *K. pneumonia* and the other test organisms were least sensitive towards the different parts of *V. rosea*.

The higher antioxidant and moderate antimicrobial activity may be due to the presence of phytochemicals like phenolic compounds, flavonoids, coumarin, quinones etc., as proved by the phytochemical analysis (Table 5).

Conclusion

The present study revealed that the all the parts of *Vinca rosea* showed better radical scavenging activity when compared to *Calotropis gigantea*. Within different parts of *Calotropis gigantea*, stem showed highest antioxidant activity at 800 µg when compared to its leaves, roots and flower. The results of this study also revealed that the zone of inhibition was different for different parts of *C. gigantea* and *V. rosea* against different microorganisms tested. The highest inhibition zone of 14, 14 and 11 mm was exerted by *Calotropis gigantea* against *Escherichia coli*, *Salmonella typhi* and *Shigella sonnei* when compared to *Vinca rosea*. Hence, the present study supports the view that these medicinal plants might be useful as antioxidant and antimicrobial agents. Further research should be aimed on adapting suitable methods so that the active principles are isolated and at the same time its efficacy is preserved during extraction and concentration.

Acknowledgements

The authors are grateful to UGC, New Delhi for providing financial support to carry out this project and to the Dept. of Microbiology, Auxilium College for their support in the microbial studies.

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